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## ***Cercospora Beticola* Toxins. Part XI: Isolation and Structure of Beticolin 0.**

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*Abstract* : Structure of beticolin 0 has been established through examination of its spectroscopic data including X-Ray diffraction and through its chemical transformation into the previously identified beticolin 2.

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We have isolated from the mycelial extract of a *Cercospora Beticola* strain a number of yellow compounds, one of which was previously known as CBT (*Cercospora Beticola* toxin)<sup>2</sup>. Structure elucidation of some of them showed that they have closely related structures<sup>3-7</sup>. These compounds can be divided in sub-groups according to i) the ring closure of the heterocycle, which can occur with the oxygen being in *para* position of the chlorine atom (*p*-beticolin) or with the oxygen being in the *ortho* position (*o*-beticolin), ii) the relative configuration at C-2 and C-3. Indeed, we observed that in the *o*-beticolin sub-group, substituents on C-2 and C-3 can be either on different sides of the mean plan of the cycle A (*o*-beticolin) or on the same side of this plan (*epi-o*-beticolin). We also showed that the *o*-beticolin skeleton can be transformed under basic conditions in the thermodynamically more stable *p*-beticolin skeleton through the opening of the heterocycle<sup>7</sup>, the minor products of this transformation being *epi-o*-beticolins (beticolin 6 and beticolin 8 from beticolin 2 and 4 respectively).

In our previous papers we tried to find spectroscopic and chemical criteria to differentiate *o*- and *p*-beticolins. First, *p*-beticolins (beticolin 1 and 3) were shown to form complexes with divalent cations such as magnesium to afford stable symmetric dimeric compounds with two magnesium atoms<sup>8</sup>, each of them being attached to the conjugated keto-enol system of the xanthone of one molecule and to the anthraquinone moiety of the other one<sup>6</sup>. Second, the easy transformation in basic conditions of the *o*-beticolins into the corresponding *p*-beticolins<sup>7</sup>, when *p*-beticolins remained unchanged in the same conditions, should allow a rapid discrimination between both skeletons. Third, NOESY experiments allowed observation of characteristic nOe's for both forms<sup>6,7</sup>.



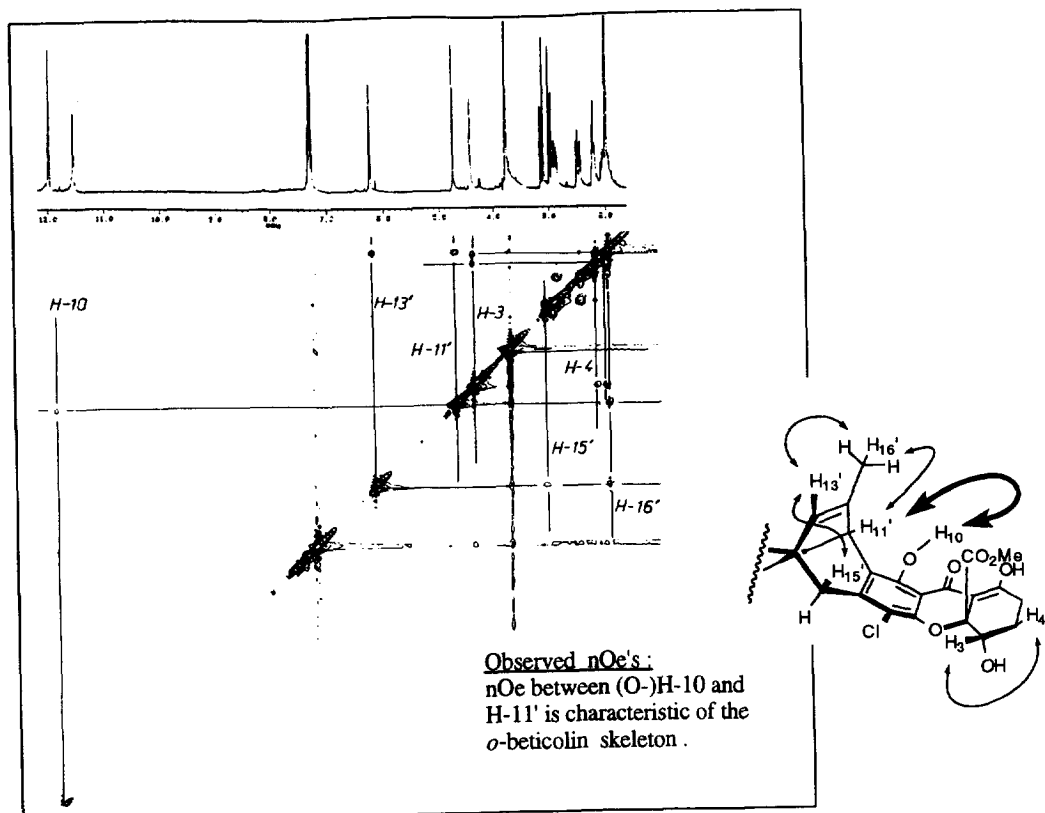


Fig 1: phase sensitive NOESY of beticolin 0 (400 MHz,  $\text{CDCl}_3$ , mixing time 0.9 s, 256 experiments)

Finally, monocrystals were obtained and X-ray diffraction analysis confirmed the structure of beticolin 0 as shown in Fig. 210.

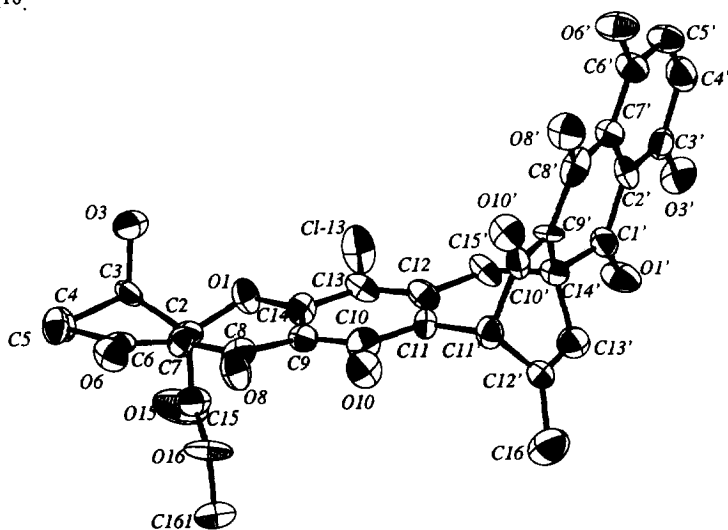


Fig.2 : ORTEP view of beticolin 0 (ellipsoids are drawn at the 50% probability level)

These last results demonstrated the validity of the chemical and spectroscopic criteria which were developed in our previous papers to differentiate *p*- from *o*-beticolins. Furthermore the *o*-beticolins seem to give mono crystals more easily than the corresponding *p*-beticolins<sup>3,5</sup>. These results will help us in structure elucidation of the minor compounds present in crude mycelial extracts.

## References and notes

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- 9- After extraction with ethyl acetate from *C. beticola* mycelium, beticolins were separated by flash chromatography using silica gel pretreated with Ca(H<sub>2</sub>PO<sub>4</sub>)<sub>2</sub>.H<sub>2</sub>O and H<sub>3</sub>PO<sub>4</sub> (Balis, C.; Payne, M. G. *Phytopathology* **1971**, *61*, 1477) and eluted with CHCl<sub>3</sub>. TLC analysis was performed using two systems : 1) CHCl<sub>3</sub>/MeOH/CH<sub>3</sub>COOH 100/2/1 and 2) Hexane/ethyl acetate 1/1. Crystallization from ethyl acetate/hexane afforded **Beticolin 0**: Rf : 0.49 (system1), 0.38 (system 2), mp = 222-228°C, [α]<sub>D</sub> = +528 (c = 0.9, CH<sub>2</sub>Cl<sub>2</sub>).  
<sup>1</sup>H NMR δ (ppm) CD<sub>3</sub>COCD<sub>3</sub> (400 MHz) : 14.9 (s, 1H), 13.95 (s, 1H), 12.55 (s, 1H), 12.0 (s, 1H), 11.58 (s, 1H), 7.47, 7.50 (2H J=10 Hz, H-4', H-5'), 6.37 (bs, 1H, H-13'), 4.83 (bs, 1H, H-11'), 4.56 (dd, 1H, J=4, 2.5 Hz, H-3), 3.81 (s, 3H, H-16), 3.12, 3.32 ( 2H, J=18 Hz H-15'), 2.95 (ddd, 1H, J=17, 11, 7 Hz, H-5b), 2.59 (ddd, 1H, J=19, 7, 1 Hz, H-5a), 2.15-2.23 (m, 2H, H-4), 1.95 (s, 3H, H16'). Attribution of the phenolic protons is achieved through examination of the long range <sup>1</sup>H-<sup>13</sup>C coupling constants.  
<sup>13</sup>C NMR δ (ppm) (100.57 MHz): 83.83 (C-2), 64.7 (C-3), 22.26 (C-4), 23.15 (C-5), 179.77 (C-6), 99.48 (C-7), 186.08 (C-8), 105.76 (C-9), 155.05 (C-10), 115.24 (C-11), 142.27 (C-12), 113.04 (C-13), 153.40 (C-14), 169.24 (C-15), 51.6 (C-16), 200.68 (C-1'), 112.66 (C-2'), 151.5 (C-3'), 123.99 (C-4'), 127.7 (C-5'), 155.79 (C-6'), 111.19 (C-7'), 180.8 (C-8'), 103.91 (C-9'), 185.4 (C-10'), 43.3 (C-11'), 140.4 (C-12'), 125.62 (C-13'), 41.51 (C-14'), 37.86 (C-15'), 18.40 (C-16')
- 10- Bright yellow monocrystals were obtained by slow evaporation at room temp. of a 2:1 methanol/anisole solution. The crystal parameters are as follows : Space group P2<sub>1</sub>2<sub>1</sub>2<sub>1</sub>, (Z=4) with a=20.185(6) , b=19.471(5) and c=8.236(3) Å. The structure was solved by direct methods and anisotropically refined to R = 7.9% (using 2351 observed structure factors). Details of the beticolin **0** and the uncomplexed beticolin **1** structures will be published elsewhere.

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